I. AMENDMENTS

A. In the Specification:

Please amend the paragraph bridging pages 88-89 of the specification to read as follows:

A1

Monoclonal antibodies can be isolated and purified from hybridoma cultures by a variety of well established techniques, including, for example, affinity chromatography with Protein-A SEPHAROSE gel, size exclusion chromatography, and ion exchange chromatography (Coligan et al., *supra*, 1992, see sections 2.7.1-2.7.12 and sections 2.9.1-2.9.3; see, also, Barnes et al., "Purification of Immunoglobulin G (IgG)," in Meth. Molec. Biol. 10:79-104 (Humana Press 1992), which is incorporated herein by reference). Methods of *in vitro* and *in vivo* multiplication of monoclonal antibodies is well known to those skilled in the art. Multiplication *in vitro* can be carried out in suitable culture media such as Dulbecco's Modified Eagle Medium or RPMI 1640 medium, optionally replenished by a mammalian serum such as fetal calf serum or trace elements and growth sustaining supplements such as normal mouse peritoneal exudate cells, spleen cells, bone marrow macrophages. Production *in vitro* provides relatively pure antibody preparations and allows scale-up to yield large amounts of the desired antibodies. Large scale hybridoma cultivation can be carried out by homogenous suspension culture in an airlift reactor, in a continuous stirrer reactor, or in immobilized or entrapped cell culture. Multiplication *in vivo* can be carried out by injecting cell clones into mammals histocompatible with the parent cells, for example, syngeneic mice, to cause growth of antibody producing tumors. Optionally, the animals are primed with a hydrocarbon, especially oils such as pristane (tetramethylpentadecane) prior to injection. After one to three weeks, the desired monoclonal antibody is recovered from the body fluid of the animal.

Please amend the specification at page 103, lines 6-16, to read as follows:

In order to elucidate the biological activity of myostatin, large quantities of myostatin protein were purified for bioassays. Stable Chinese hamster ovary (CHO) cell lines producing high levels of myostatin protein were generated by co-amplifying a myostatin expression cassette with a

a2 dihydrofolate reductase cassette using a methotrexate selection scheme (McPherron et al., *supra*, 1997). Myostatin was purified from the conditioned medium of the highest producing line by successive fractionation on hydroxyapatite, lentil lectin SEPHAROSE gel, DEAE agarose, and heparin SEPHAROSE gel. Silver stain analysis revealed that the purified protein obtained following these four column chromatography steps (referred to as "heparin eluate") consisted of two species with molecular masses of approximately 35 kilodaltons (kDa) and 12 kDa.

B. In the Claims

Please amend claim 16 to read as follows:

a3 16. (Amended) The peptide of claim 8, wherein the peptide is a mature myostatin comprising about amino acid residues 268 to 374 of a promyostatin polypeptide, or a functional portion thereof.